

## Hematology analyzers

## New applications, middleware, and more for hematology analyzers

Brendan Dabkowski

Sysmex WAM Management Reports module and, soon, the WAM Select. Abbott's updated Cell-Dyn Sapphire. The DxH Slidemaker Stainer from Beckman Coulter. Just some of what's new in hematology as 2011 comes to a close and the new year gets underway.

With core labs, data management, workflow, and more in mind, hematology laboratories know what they need and companies do, too. "The market is looking for solutions that impact the entire testing process to achieve efficiency, address technical labor shortages, and reduce manual differential reviews and total cost of testing," says Bill Bailey, marketing manager, hematology, Abbott Diagnostics.

To this end, says Bailey, Abbott continues to refine its Cell-Dyn Sapphire hematology analyzer and in November released an updated version of Sapphire, a system for challenging samples in high-volume laboratories. Still available, and featured with Sapphire in this month's hematology analyzers product guide, are Abbott's Cell-Dyn Ruby, to which the company added a new service pack with updated software and hardware; Cell-Dyn Emerald; and Cell-Dyn 3700. Improvements to the company's Cell-Dyn systems, along with updates to its AbbottLink remote diagnostics product, have further enhanced instrument reliability and customer satisfaction, Bailey adds.

Sysmex America will soon release the Work Area Manager (WAM) Select middleware solution for its X series automated hematology analyzers and SP-1000i slidemaker/stainer, says Sysmex's director of marketing, Alan Burton. WAM Select, which incorporates decision support software, provides an autovalidation interface to laboratory information systems and is designed for small to mid-size labs. Other products intended to enhance the company's hematology systems are the WAM Management Reports module, which provides key metrics such as turnaround times, test costs, result validation rates, and rules and result statistics; e-Supply software solution, which streamlines reagent ordering, manages customers' on-site inventory, and reduces costs and errors associated with expedited reagent shipping; and Sysmex Managed Calibration, an evidence-based calibration program that provides six-month calibration verification and as-needed calibration adjustments to X series analyzers.

New to the guide this year is CellaVision, which offers its DM96 and DM1200 digital cell morphology systems. Both systems, says vice president of sales and business development Ron Hagner, automatically locate cells on stained slides, and, using artificial neural networks, classify the white blood cells and red blood cell morphology for the operator to verify. In September, CellaVision received FDA 510(k) clearance for a body fluid application on the DM1200. Users of the application can classify nucleated cells into seven classes and share cell images and regions of interest with colleagues. The application also provides a digital scan of a sample area in 10x and 50x magnifications. Next year, says Hagner, the company will launch its Image Capture System for small labs that perform one to 15 differentials per day. The system will be a part of CellaNet, a networking solution that allows hospitals to share cell images with related labs, pathologists, and clinicians in any location.

Beckman Coulter launched last month its UniCel DxH Slidemaker Stainer analysis system, a complement to the company's DxH 800 hematology system, which automates slide making and staining, says director of hematology and hemostasis strategic marketing Ronald Hebert. Response to the DxH 800, introduced in December 2008, has been "overwhelming," says Hebert, who notes that the company has sold nearly 1,000 of the instruments. The company's HematoFlow cellular analysis solution with CytoDiff, a new product available only in Europe, combines hematology and flow cytometry with auto-gating software to characterize WBC populations.

Finally, though not yet in the product guide, the Bloodhound integrated hematology system has been getting some buzz since its debut in July at the American Association for Clinical Chemistry's annual meeting. The morphology-based analyzer consists of a digital image-based cell locator, classifier, and counter and is designed to automatically perform a CBC, five-part differential, and reticulocyte count using digital imaging. The system's viewing station, consisting of a large-screen iMac computer, allows users to manipulate images of blood cells, such as organizing cells into rows, for the purpose of studying, interpreting, or classifying the cells. "You can zoom them up; you can reclassify things; you can make interpretations a little more efficiently because you can put the cells side by side," says James Linder, MD, chief medical officer, Constitution Medical, which is developing the Bloodhound system. The company will conduct clinical trials early next year and plans to submit the results to the FDA shortly thereafter.

CAP TODAY's hematology analyzers product guide includes systems from the aforementioned companies and from Horiba Medical and Siemens Healthcare Diagnostics. Companies supplied the information listed. Readers interested in a particular system should confirm that it has the stated features and capabilities.

Brendan Dabkowski is CAP TODAY associate editor.

<b>Part 1 of 10</b>	<b>Abbott Hematology</b> Rick Gooch rick.gooch@abbott.com 5440 Patrick Henry Drive Santa Clara, CA 95054 800-933-5535 www.abbottdiagnostics.com
<b>Name of instrument</b> <b>First year installed in U.S./Outside U.S./No. of units sold in 2010</b> <b>No. units installed in U.S./Outside U.S./List price</b>	<b>CELL-DYN Sapphire*</b> 2005/2005/— >175/>750/\$250,000
<b>Test menu:</b> • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, % neut, mono, lymph, eos, baso): • Laboratory • Flags	standard menu (left) plus: MPV, RDW, retic %&#, IRF, NRBC %&#, CD61, CD3T %&#, CD4T %&#, CD8T %&#, 4/8 — band, IG, blast, variant lymph, nvWBC, rstRBC, IR, PLT clmp, ASYM, FP, CD61 agglutination, clot detected during aspiration, short sample — — — — CD61 for PLTs, CD3/4, CD3/8 (immuno T-cell)
<b>FDA-cleared tests not clinically released</b> <b>Tests not available but submitted for 510(k) clearance</b> <b>Tests in development</b> <b>Tests for research use only</b> <b>Tests unique to analyzer</b>	— — — — —
<b>Differential method(s) used</b>	<b>MAPSS (Multi-Angle Polarized Scatter Separation) and three-color fluorescence</b> 0.4–250.0 × 10 <sup>3</sup> µL/ 0.22–7.50 × 10 <sup>6</sup> µL 1.0–24.8 g/dL/11.0–2,000.0 × 10 <sup>3</sup> µL 37.0–179 fL (MCV)
<b>Linearity:</b> • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%)	≤2.7 percent/≤1.5 percent ≤1.0 percent/≤4.0 percent ≤1.0 percent (MCV)
<b>Precision:</b> • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct	—
<b>Accuracy of automated differential compared with manual differential (per CLSI H-20A)</b>	neut% r=0.942 slope 0.947 y=0.446; lym% r=0.936 slope=0.943 y=2.811; mono% r=0.623 slope=1.057 y=0.851; eos% r=0.446 slope=1.024 y=0.288; baso% r=0.232 slope=0.257 y=0.350
<b>Interfering substances:</b> • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	PLT clumps, neutrophil aggregates, HbC crystals, lyse-resistant RBCs, cryoglobulin, cryofibrinogen, fragmented WBC, NRBC autoagglutination, cold agglutinins, elevated WBC, giant PLTs, hemolysis, small WBC autoagglutination, cold agglutinins, elevated WBC, giant PLT, hemolysis, hyperglycemia auto and cold agglutination, cryoglobulin, cryofibrinogen, giant PLT, micro RBC, PLT clumps, RBC fragments, WBC fragments, PLT satellitism lipids >700 mg/dL, WBCs >250 × 10 <sup>9</sup> /L, bilirubin >33 mg/dL, HbC crystals see WBC
<b>Interfering substances: differential</b>	—
<b>Maximum CBCs per hour/Maximum CBCs and differentials per hour</b> <b>Minimum specimen volume open/Closed/Sample dead volume closed</b> <b>Microsample capability</b> <b>Prepares microscope slides automatically or flags problems for slide prep</b> <b>No. of automatic slidemakers available/List price</b>	105/105 120 µL/120 µL/0.5 mL, 0.3 mL for 10.25 × 64 mm tubes yes no —/\$125,000
<b>Archives patient data/Previous patient results included with recent results</b> <b>Maximum archived data accessible when system online</b> <b>No. specimens for which numeric results saved in memory at once</b> <b>No. specimens for which histo/cytogram results saved in memory at once</b> <b>Performs delta checks</b> <b>Tags and holds results for followup, confirmatory testing, or rerun</b> <b>Parameters for flags for holding samples defined by user or vendor</b> <b>Scattergram display: cell-specific color</b> <b>Histogram display: color with thresholds</b> <b>User interface can display choice of specimen/result information</b>	yes/yes 10,000 results 10,000 results 10,000 results yes yes user or vendor yes yes yes
<b>LIS interface formats supported</b> <b>Information transferred on LIS interface</b>	ASTM 1394 numeric and flag results, instrument to LIS; patient demographics, patient orders, LIS to instrument—broadcast; host query for patient demographics and orders
<b>LOINC codes transmitted with all results</b> <b>Interface available or planned to automated specimen-handling system</b> <b>Bar-code symbologies read on specimen tube</b> <b>Accommodates bar-code placement per CLSI standard Auto2A</b>	no none Codabar, codes 39 and 128, Interleaved 2 of 5 yes
<b>Time required for maintenance by lab personnel</b> <b>Onboard diagnostics for troubleshooting/Limited to software problems</b> <b>Manufacturer can perform diagnostics via modem</b>	daily: 30 seconds; weekly: 10 minutes; monthly: 5 minutes yes/no yes
<b>Distinguishing features (supplied by company)</b>	four optical and three fluorescent detectors provide multiple scatterplot analysis; 2-D optical platelets prevent interferences; fluorescent analysis of reticulocytes, NRBCs, and three-color monoclonal analysis on routine hematology analyzer; OpenFlow MAb test selections

Note: a dash in lieu of an answer means company did not answer question or question is not applicable

\*please see the CELL-DYN Sapphire operator's manual for product labeling, including warnings, limitations, and precautions

## Hematology analyzers

Part 2 of 10	Abbott Hematology Rick Gooch rick.gooch@abbott.com 5440 Patrick Henry Drive Santa Clara, CA 95054 800-933-5535 www.abbottdiagnostics.com	Abbott Hematology Karen Busch karen.busch@abbott.com 5440 Patrick Henry Drive Santa Clara, CA 95054 800-933-5535 www.abbottdiagnostics.com	Abbott Hematology Rick Gooch rick.gooch@abbott.com 5440 Patrick Henry Drive Santa Clara, CA 95054 800-933-5535 www.abbottdiagnostics.com
Name of instrument First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	CELL-DYN Ruby* 2006/2006/— >450/>1,500/\$185,000	CELL-DYN Emerald* 2009/2008/0 >950/>1,600/\$30,000	CELL-DYN 3700* 1999/1999/— >200/>1,500/\$180,000 SL Model, \$140,000 CS model
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, % neut, mono, lymph, eos, baso):  • Laboratory • Flags  FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only Tests unique to analyzer	standard menu (left) plus: MPV, RDW, retic #& percent  — NRBC, FWBC, NWBC, RRBC, band, IG, blast, variant lymph, RBC morph., DFLT, MCHC, LRI, URI, LURI, ATYPDEP, high/low interp. message, WBC  — — — — atypical depolarization flag	WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, lymph percent&#, gran percent&#, mid percent&#, RDW, MPV  — dispersional data alerts, suspect measurand flags and count invalidation flags  — — — — none	standard menu (left) plus: RDW, MPV, retic #&%, IRF  — suspect populations, band, blast, variant lymph, IG, NRBC, RRBC, NWBC, LRI, URI, LURI, RBC morphology, FWBC, high/low interpretation message, WBC  — — — — IRF, veterinary capabilities**
Differential method(s) used	MAPSS (Multi-Angle Polarized Scatter Separation)	impedance counting	MAPSS (Multi-Angle Polarized Scatter Separation)
Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%)	0.02–246 × 10 <sup>3</sup> /μL/0.00–7.50 × 10 <sup>6</sup> /μL 0.00–25.0 g/dL/0.00–3,000 × 10 <sup>3</sup> /μL 58–139 fL (MCV)	0.4–96.1 K/μL/0.22–7.61 M/μL 3.3–24.6 g/dL/9–1,375 K/uL 5.3–75.6 percent (Hct)/48.8–115 fL (MCV)	0–250 K/μL/0–8 M/μL 0–24 g/dL/0–2,000 K/μL 50–200 fL (MCV)
Precision: • WBC count/RBC count  • Hemoglobin/platelet  • MCV or Hct	2.4 percent/1.8 percent  1.4 percent/3.8 percent  0.8 percent (MCV)	3.5 percent (95 percent confidence limit)/2.0 percent (95 percent confidence limit) 2.1 percent (95 percent confidence limit)/6.1 percent (95 percent confidence limit) 1.7 percent Hct (95 percent confidence limit)/0.8 percent MCV (95 percent confidence limit)	≤2.5 percent/≤1.5 percent  ≤1.2 percent/≤5.0 percent  ≤1.0 percent (MCV)
Accuracy of automated differential compared with manual differential (per CLSI H-20A)	neut percent r=0.983, slope=0.97, y=-1.98; lymph r=0.921, slope=0.95, y=0.94; mono r=0.711, slope=1.10, y=1.93; eos r=0.952, slope=1.04, y=0.01; baso r=0.146, slope=0.18, y=1.22	—	neut #&%= ≥0.95, lymph #&%= ≥0.94, mono #&%= ≥0.86, eos #&%= ≥0.84, baso #&%= ≥0.73
Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	fragile WBC, neutrophil aggregates, lytic-resistant RBC, NRBC, PLT clumps, cryofibrinogen, cryoglobulin  elevated WBC, increased numbers of giant PLT, autoagglutination, in vitro hemolysis  MCV: elevated WBC, hyperglycemia, in vitro hemolysis, increased number of giant PLTs  WBC fragments, in vitro hemolysis, microcytic RBC, cryofibrinogen, cryoglobulins, PLT clumping, increased number of giant PLT  elevated WBC, increased plasma substances (triglycerides, bilirubin, in vivo hemolysis), lytic-resistant RBC	cryoglobulin, cryofibrinogen, heparin, monoclonal proteins, nucleated red cells, platelet clumping, unlysed red cells, clotting, smudge cells, uremia plus immunosuppressants cryoglobulin, cryofibrinogen, giant platelets, high white cell count (>50,000 K/μL), autoagglutination, clotting, hemolysis (in vitro), microcytic red cells cryoglobulin, cryofibrinogen, giant platelets, high white cell count (>50,000 K/μL) hyperglycemia (>600 mg/dL), autoagglutination, clotting, hemolysis (in vitro), microcytic red cells, reduced red cell deformability, swollen red cells cryoglobulin, cryofibrinogen, hemolysis (in vivo and in vitro), microcytic red cells, red cell inclusions, white cell fragments, clotting, giant platelets, heparin, platelet clumping, platelet satellitosis carboxyhemoglobin (>10 percent), cryoglobulin, cryofibrinogen, hemolysis (in vivo) heparin, high white cell count (>50,000 K/μL), hyperbilirubinemia, lipemia, monoclonal proteins platelet aggregates, NRBCs, giant platelets, cryoglobulins, incomplete lysis of RBC, small lymphocytes, fibrin clots, shift in WBC cell distribution due to EDTA anticoagulant equilibration	NRBCs (WIC only), lytic-resistant RBCs, PLT clumps, cryoglobulin and cryofibrinogen, fragile WBCs  increased number giant PLTs, autoagglutination, in vitro hemolysis  MCV: elevated WBC count, increased number giant PLTs, hyperglycemia, in vitro hemolysis  WBC fragments, in vitro hemolysis, microcytic RBCs, cryoglobulin, PLT clumps, increased number giant PLTs  increased plasma substances (triglycerides, bilirubin, in vivo hemolysis), lyse-resistant RBCs
Interfering substances: differential	fragile WBC, neutrophil aggregates, lytic-resistant RBC, NRBC, PLT clumps, cryofibrinogen, cryoglobulin, paraproteins	—	see WBC
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed	84/84 150 μL/230 μL/1.2 mL	60/60 9.8 μL/—/—	90/90 130 μL/355 μL/1.0 mL
Microsample capability Prepares microscope slides automatically or flags problems for slide prep No. of automatic slidemakers available/List price	no no —/\$125,000	no no —	yes yes (flags only) —/\$125,000
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes/yes 10,000 results 10,000 results 10,000 results no yes user or vendor yes yes yes	yes/no 60,000 on USB and 1,500 results on internal memory 60,000 on USB and 1,500 results on internal memory 60,000 on USB and 1,500 results on internal memory no no no no no yes	yes/yes 10,000 results 10,000 results 10,000 results no yes user or vendor yes yes yes
LIS interface formats supported Information transferred on LIS interface  LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube	LIS1/LIS2 CLSI numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, patient orders, LIS to instrument—broadcast; host query for patient demographics and orders  no — Codabar, codes 39 and 128, Interleaved 2 of 5, ISBT	proprietary (instrument or vendor specific) numeric and flag results, instrument to LIS  no — Codabar, codes 39 and 128, Interleaved 2 of 5, Chinese post, code 93, EAN8, EAN13, EAN128, IATA, industrial 2 of 5, Italian pharmaceutical, matrix 2 of 5, MSI/Plessey, UK/Plessey, Telepen, TriOptic, S-Code, UPC A, UPC E	proprietary numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast  no — Codabar, codes 39 and 128, interleaved 2 of 5
Accommodates bar-code placement per CLSI standard Auto2A	yes	yes	yes
Time required for maintenance by lab personnel  Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	daily: 30 seconds; weekly: 5 minutes; monthly: 10 minutes yes/no yes	daily: 3 minutes; monthly: 5 minutes; bi-annually: 10 minutes no/no no	daily: 30 seconds; bi-weekly: 5 minutes; monthly: 10 minutes yes/no —
Distinguishing features (supplied by company)	touch-sensitive screen, all optical technology; onboard maintenance videos; lyse-resistant RBC mode; rules-based result annotations	small: sample size, reagent volumes used, and physical size; reliable: system averages one service call per year; easy to use: system has touchscreen software with intuitive icons and minimal layers	MAPSS cell-by-cell analysis; reticulocyte with reportable IRF (immature reticulocyte fraction); up to 60 different animal types can be configured for analysis  <small>*please see the CELL-DYN 3700 operator's manual for product labeling, including warnings, limitations, and precautions **veterinary applications for medical devices are not currently subject to premarket regulation by FDA</small>
<small>Note: a dash in lieu of an answer means company did not answer question or question is not applicable</small>	<small>*please see the CELL-DYN Ruby operator's manual for product labeling, including warnings, limitations, and precautions</small>	<small>*please see the CELL-DYN Emerald operator's manual for product labeling, including warnings, limitations, and precautions</small>	

## Hematology analyzers

Part 3 of 10	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Blvd Brea, CA 92821 305-380-3060 www.beckmancoulter.com	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Blvd Brea, CA 92821 305-380-3060 www.beckmancoulter.com	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Blvd Brea, CA 92821 305-380-3060 www.beckmancoulter.com
Name of instrument First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	UniCel DxH 800 2008/2008/493 (Sept. 2011 YTD) 540/371/\$229,000	LH 1500 Hematology Automation Series 2002/2003/15 (Sept. 2011 YTD) >65/25/varies	LH 780/LH 785 2006/2007/170 (Sept. 2011 YTD) 629/595/LH 780: \$214,500
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, % neut, mono, lymph, eos, baso):  • Laboratory • Flags  FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only  Tests unique to analyzer	standard menu (left) plus: RDW-CV, RDW-SD, MPV, retic#, retic%, IRF, MRV, NRBC# and %, body fluids-total nucleated count, and RBC count for synovial, serous, and CSF fluids — definitive, suspect and system messages, user-definable extended decision rules, ISLH consensus rules, user-definable differential sensitivity — — high light scatter reticulocytes (HLR% and HLR#), low hemoglobin density (LHD), microcytic anemia factor (MAF), mean spheroid cell volume (MSCV), plateletcrit (PCT), platelet distribution width (PDW), reticulocyte distribution width (RDWR-CV and RDWR-SD), red cell size factor (RSF), cell population data research parameters	standard menu (left) plus: RDW, MPV, retic %&#, IRF, graded RBC morph., NRBC %&#, TNC & RBC on CSF, synovial, and serous fluids — user-definable age-, gender-, and/or location-based reference intervals; action and critical limits; user-definable RBC morphology; user-selectable sensitivity for differential, abnormal population suspect messages — — MSCV, HLR %&#, PDW, PCT, WBC research population data (RPD) LH 780: MAF, RSF, RDWR-SD, RDWR-CV  IVD: NRBC, body fluids; RUO: MSCV, WBC RPD	standard menu (left) plus: RDW, RDW-SD, MPV, retic %&#, IRF, MRV, graded RBC morph., NRBC %&#, TNC and RBC on CSF, synovial, and serous fluids — user-definable age-, gender-, and/or location-based reference intervals; action and critical limits; user-definable RBC morphology; user-definable sensitivity for differential abnormal populations, suspect and definitive messages — — — RSF, MAF, MSCV, HLR %&#, RDWR-CV, RDWR-SD, PDW, PCT, WBC research population data (RPD)  IVD: NRBC, body fluids, RDW-SD; RUO: MSCV, RSF, MAF, WBC RPD
Differential method(s) used  Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%) Precision: • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct Accuracy of automated differential compared with manual differential (per CLSI H-20A)  Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin  Interfering substances: differential	flow cytometric digital analysis using volume, conductivity, and five angles of light scatter, digital signal processing, advanced algorithm applications, high-definition cellular resolution, DataFusion 0-400/0-8.5 0-25.5/0-3,000 50-150 (MCV) ≤3.0 percent/≤1.5 percent ≤1.5 percent/≤3.5 percent ≤1.0 percent NE = ±2.0; LY, MO = ±3.0; EO, BA = ±1.0 (or 10% percent, whichever is greater)  precipitated elevated proteins, cryoglobulin, fragmented white cells, agglutinated white cells, lyse-resistant red cells, giant platelets, platelet clumps, unlysed particles >35 fL in size very high WBC count, high concentration of very large platelets, autoagglutination very high WBC count, high concentration of very large platelets, autoagglutination platelet clumps, white cell fragments, very small red cells, red cell fragments, giant platelets, electric noise severe lipemia, heparin, certain unusual RBC abnormalities that resist lysing  elevated triglycerides, precipitated elevated proteins	Coulter's 3-D VCS biophysical flow cytometry with IntelliKinetics, AccuGate, and AccuFlex technologies  0-400/0-8.0 0-25/0-3,000 50-200 (MCV) <1.7 percent/<0.8 percent <0.8 percent/<3.3 percent <0.8 percent (MCV) lymph% = ±3.0%, —; neut% = ±3.0%, —; mono% = ±2.0%, eos% = ±1.0%, baso% = ±1.0%  unusual RBC abnormalities that resist lysing, NRBC, fragmented WBC, unlysed particle >35 fL, giant PLT, PLT clumps  very high WBC, high concentration large PLT, autoagglutinins very high WBC, high concentration large PLT, autoagglutinins very small RBCs and WBC fragments may interfere  very high WBC, severe lipemia, heparin, rare lyse-resistant RBCs  high triglycerides may affect lysing	Coulter's 3-D VCS biophysical flow cytometry with IntelliKinetics, AccuGate, and AccuFlex technologies  0-400/0-8.0 0-25/0-3,000 — ≤1.7 percent/≤0.8 percent ≤0.8 percent/≤3.3 percent ≤0.8 percent (MCV) lymph% = ±3.0%, neut% = ±2.0%, mono% = ±3.0%, eos% = ±1.0%, baso% = ±1.0%  unusual RBC abnormalities that resist lysing, NRBC, fragmented WBC, unlysed particle >35 fL, giant PLT, PLT clumps  very high WBC, high concentration large PLT, autoagglutinins very high WBC, high concentration large PLT, autoagglutinins (MCV) very small RBCs and WBC fragments  very high WBC, severe lipemia, heparin, rare lyse-resistant RBCs  high triglycerides may affect lysing
Maximum CBCs per hour/Maximum CBCs and differentials per hour  Minimum specimen volume open/Closed/Sample dead volume closed Microsample capability Prepares microscope slides automatically or flags problems for slide prep No. of automatic slidemakers available/List price	>100 per hour/>100 per hour  165 µL/165 µL/300-400 µL yes yes —/DxH SMS \$165,000	110 per analyzer on automation system/110 per analyzer on automation system 200 µL/300 µL, 550 µL with slidemaker/1.0 mL yes yes >850 (U.S.)/\$110,000	110/110; 105/100 with SMS  200 µL/300 µL (550 µL with slidemaker)/1.0 mL yes yes >500/\$110,000
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes/yes 40,000 standalone 40,000 standalone 40,000 yes yes yes yes yes yes	yes/yes 20,000 samples per instrument 20,000 samples per instrument 20,000 samples per instrument yes yes user or vendor yes yes yes	yes/yes 20,000 samples 20,000 samples 20,000 samples yes yes user or vendor yes yes yes
LIS interface formats supported Information transferred on LIS interface  LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube Accommodates bar-code placement per CLSI standard Auto2A	CLSI LIS1-A numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, patient orders, LIS to instrument—broadcast; host query for patient demographics and orders (available with release of workcell) yes Beckman Coulter Codabar, codes 39 and 128, interleaved 2 of 5, NW7 yes	— numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, patient orders, LIS to instrument—broadcast  no Beckman Coulter Codabar, codes 39 and 128, interleaved 2 of 5, NW7 yes	proprietary numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, patient orders, LIS to instrument—broadcast  no Beckman Coulter Codabar, codes 39 and 128, interleaved 2 of 5, NW7 yes
Time required for maintenance by lab personnel  Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	no routine maintenance, all maintenance procedures are on as-needed basis  yes/no yes	daily, weekly, monthly, and as needed maintenance procedures, however time varies by automation line  yes/no yes	no routine maintenance; only as needed  yes/no yes
Distinguishing features (supplied by company)	integrated automation w/auto repeat/reflex testing based on extended onboard user-defined decision rules; single aspiration pathway negates mode-to-mode comparisons; flow cytometric digital morphology w/five angles of light scatter; separate channel for WBC, NRBC, and reticulocyte analysis; digital signal processing, DataFusion; future scalability	system automatically loads and unloads cassettes, performs reflex and repeat testing, sorts tubes for off-line tests, stores tubes with availability for retrieval for any test type; multiple configurations available; RUO: WBC research population data	extensive onboard user-defined decision support; extended linearity for WBC and PLT using AccuCount technology; enumeration of NRBCs with every differential; random access/automation ready; integrated slidemaker/slidestainer options; proservice; electronic IQAP; expanded QC module; RUO: WBC research population data

Note: a dash in lieu of an answer means company did not answer question or question is not applicable

## Hematology analyzers

Part 4 of 10	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Boulevard Brea, CA 92821 305-380-3060 www.beckmancoulter.com	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Boulevard Brea, CA 92821 305-380-3060 www.beckmancoulter.com	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Boulevard Brea, CA 92821 305-380-3060 www.beckmancoulter.com
Name of instrument First year installed in U.S./Outside U.S./No. of units sold in 2010	Coulter LH 750 2001/2001/261 (Sept. 2011 YTD)	Coulter LH 500 2003/2003/198 (Sept. 2011 YTD)	Coulter HmX 1999 HmX AL/—/209 (Sept. 2011 YTD)
No. units installed in U.S./Outside U.S./List price	2,065/1,473/\$195,000	1,360/956/\$145,000	>1,232/>1,213/\$135,000
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, mono, lymph, eos, baso):  • Laboratory • Flags  FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only  Tests unique to analyzer	standard menu (left) plus: RDW, MPV, retic #&%, IRF, MRV, graded RBC morph., NRBC %&#, TNC & RBC on CSF, synovial, and serous fluids — user-definable age-, gender-, and/or location-based reference intervals; action and critical limits; user-definable RBC morphology; gradient messages (=+, ++, +++); user-selectable sensitivity for differential abnormal population suspect messages — MSCV, HLR %&#, PDW, PCT, WBC research population data (RPD) IVD: NRBC, body fluids; RUO: MSCV, WBC RPD	standard menu (left) plus: retic #, retic %, MRV, IRF, RDW, MPV — user-definable age-, gender- and/or location-based reference intervals, action and critical limits; user-definable RBC morphology; gradient messages — PCT, PDW, WBC RPD	standard menu (left) plus: RDW, MPV, retic #&%, graded RBC morph., IRF, MRV — comprehensive high/low, definitive and suspect messages — PCT, PDW
Differential method(s) used	Coulter's 3-D VCS biophysical flow cytometry with IntelliKinetics, AccuGate, and AccuFlex technologies	Coulter's 3-D biophysical flow cytometry with AccuGate 500, Reaction Manager technologies	Coulter's 3-D VCS technology
Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%)	0–400/0–8.0 0–25/0–3,000 —	0–200/0–7.0 0–25/0–2,000 50–150 (MCV)	0–99.9/0–7.0 0–25/0–999 50–150 (MCV)
Precision: • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct	≤1.7 percent/≤0.8 percent ≤0.8 percent/≤3.3 percent ≤0.8 percent (MCV)	≤2.5 percent/≤2.0 percent ≤1.5 percent/≤5.0 percent ≤2 percent (MCV)	<2.5 percent/<2.0 percent <1.5 percent/<5.0 percent <2.0 percent (MCV)
Accuracy of automated differential compared with manual differential (per CLSI H-20A)	lymph% = ±3.0%, neut% = ±2.0%, mono% = ±3.0%, eos% = ±1.0%, baso% = ±1.0%	lymph= ±1.5 % mean diff., mono= ±1.5 % mean diff., neut= ±2.0% mean diff., eos= ±0.5 % mean diff., baso= ±0.5 % mean differential	lymph%= ±3.0%, —; mono%= ±2.0%, —; neut%= ±3.0%, —; eos%= ±1.0%, —; baso%= ±1.0%, —
Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	unusual RBC abnormalities that resist lysing, NRBC, fragmented WBC, unlysed particle >35 fL, giant PLT, PLT clumps very high WBC, high concentration large PLT, autoagglutinins  MCV and Hct: very high WBC, high concentration large PLT, autoagglutinins  very small RBCs and WBC fragments may interfere  very high WBC, severe lipemia, heparin, rare	lyse-resistant, nucleated RBCs, frag. WBCs, agglutination WBCs, unlysed particles >35 fL, very large or agg. PLTs, fibrin, cell frag., or other debris very high WBC count, many very large PLTs, agglutinin RBCs, RBCs <36 fL, fibrin, cell fragments, or other debris MCV: very high WBC count, high concentration of very large PLTs, agglutinin RBCs, RBC fragments <36 fL, rigid RBCs very small red cells near the upper threshold, cell fragments, clumped PLTs, PLT fragments or cellular debris near the lower PLT threshold, giant PLTs, PLT clumps, red and white cell fragments, electronic noise, very small red cells very high WBC count, severe lipemia, heparin, lyse-resistant RBCs, turbidity such as elevated triglycerides	unusual RBC abnormalities that resist lysing, NRBC, fragmented WBC, unlysed particle >35 fL, large PLT  very high WBC, high concentration of very large PLT, autoagglutinins  MCV and Hct: very high WBC, high concentration of large PLT, autoagglutinins  very small RBCs and WBC fragments may cause no fit  very high WBC, severe lipemia, heparin, rare lyse-resistant RBCs
Interfering substances: differential	lyse-resistant RBCs high triglycerides may affect lysing	factors that affect WBC count above or high triglycerides that affect lysing, hypogran. anuloocytes, agranul. granulocytes, lyse-resist. red cells, very small or multi-population lymphocytes, elevat. trigly., precipitated elev. proteins	high triglycerides may affect lysing
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed Microsample capability Prepares microscope slides automatically or flags problems for slide prep	110/110 200 µL/300 µL, 550 µL with slidemaker/1.0 mL yes yes, both	75/70 125 µL/185 µL/tube dependent yes no	75/75 125 µL/185 µL/50 µL predilute/0.5 mL yes no
No. of automatic slidemakers available/List price	>1,000 (U.S.)/\$110,000	—	—
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes/yes 20,000 samples 20,000 samples 20,000 samples yes yes user or vendor yes yes yes	yes/yes 20,000 samples 20,000 samples 20,000 samples yes yes user yes yes yes	yes 5,000 samples 5,000 samples 5,000 samples no yes user or vendor four colors/cell types colors without thresholds no
LIS interface formats supported Information transferred on LIS interface  LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube  Accommodates bar-code placement per CLSI standard Auto2A	RS-232, proprietary numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast no Beckman Coulter Codabar, codes 39 and 128, Interleaved 2 of 5, NW7 yes	RS-232, proprietary numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast no — Codabar, codes 39 and 128, Interleaved 2 of 5, NW7 yes	RS-232, proprietary numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast no — Codabar, codes 39 and 128, Interleaved 2 of 5, NW7 no
Time required for maintenance by lab personnel	no routine maintenance; only as needed	no routine maintenance; only as needed	none
Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	yes/no yes	yes/no yes	yes/no no
Distinguishing features (supplied by company)	extensive decision support; enumeration of NRBCs with every differential; random access; automation ready; extended linearity for WBC and PLTs; RUO: WBC RPD	extensive decision support, extended linearity for WBC and PLT, low review rate, small footprint, superior reliability, ProService, electronic IQAP	VCS technology; low review rate; no routine daily maintenance; triplicate counting; aperture burn circuit; sweepflow; SmartStart system; autoloader and single-sample models

Note: a dash in lieu of an answer means company did not answer question or question is not applicable

## Hematology analyzers

Part 5 of 10	Beckman Coulter Hamid Erfanian herfanian@beckman.com 250 South Kraemer Boulevard Brea, CA 92821 305-380-3060 www.beckmancoulter.com	CellaVision Ron Hagner ron.hagner@cellavision.com 4107 Burns Road Beach Gardens, FL 33410 919-619-3909 www.cellavision.com	HORIBA Medical Jim Knowles jimknowles@horiba.com 34 Bunsen Irvine, CA 92618 888-903-5001 ext. 4553 www.horiba.com/us/en/medical
Name of instrument First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	Coulter Ac•T 5diff Family; Ac•T 5diff AL 2001/2000; 2003/2003; 317 (combined Sept. 2011 YTD) 1,029/1,454 combined in and out US \$38,500 (CP)/\$4,500 (AL)	CellaVision DM96 and CellaVision DM1200 2004/2003/— ~400/~700/~\$135,000–\$175,000	Pentra 60C+ Hematology Analyzer 2000/2000/85 >350/>600/\$47,313
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, mono, lymph, eos, baso):  • Laboratory  • Flags FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only  Tests unique to analyzer	standard menu (left) plus: RDW, MPV  atypical lymphocytes # (ATL#), atypical lymphocytes % (ATL%), immature cells # (IMM#), immature cells % (IMM%), PCT, PDW complete operator-selectable flagging — — — PCT, PDW, IMM, ATL	%&# neut, mono, lymph, eos, baso, segmented, bands, blast, promyelocytes, myelocytes, metamyelocytes, variant lymphocytes, plasma cells, giant platelets, platelet clumps, erythroblasts; RBC morphology pre-characterizations include anisocytosis, poikilocytosis, polychromasia, microcytosis, macrocytosis, hypochromia — — — — analysis of cytocentrifuged samples, body fluids (reported parameters: neutrophils, eosinophils, lymphocytes, macrophages (including monocytes), other (basophils, lymphoma cells, atypical lymphocytes, blast cells, and tumor cells)	standard menu (left) plus: RDW, MPV  atypical lymphocytes, atypical lymphocytes %, LIC, LIC %  operator-selectable flagging — — — PCT, PDW, ATL, LIC
Differential method(s) used	A•V technology combining cytochemistry, focused flow impedance, and light absorbance principles of measurement	light microscopy, image analysis, and artificial neural networks	DHSS technology combining cytochemistry, focused flow impedance, and light absorbance principles of measurement
Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%)	0.4–90.0/23–7.7; AL: 0.4–120.0/0.3–8.0 0–22.9/4–1000; AL: 1.3–24.0/10.0–1,000 1.8–63.8 (Hct)	— — —	0–120/0–8 0–24/0–1,900 0–67 (Hct)
Precision: • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct	<2 percent/<2 percent <1 percent/<5 percent <2.0 percent (Hct); AL: <2.0% (Hct)	— — —	<2 percent/<2 percent <1 percent/<5 percent <2 percent (Hct)
Accuracy of automated differential compared with manual differential (per CLSI H-20A)	not available in CLSI H-20A format	seg neut% y=0.97x + 1.3 r= 0.987, lymph% y=0.97x + 1.2 r= 0.979, eos% y=1.01+0.1, r=0.960, mono% y=0.97+0.2, r=0.941, band neut% y=0.87x+0.1, r=0.917	neut% r=0.99, —; lymph% r=0.98, —; mono% r=0.96, —; eos% r=0.89, —; baso% r=0.54, —
Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	NRBCs, PLT clumps, large PLTs, lyse-resistant RBCs  cold agglutinins, PLT clumps, WBC overlinearity  Hct: lipemic samples, high WBC, cold agglutinins  RBC and WBC fragments  elevated WBC, lipemia	— — — — —	NRBCs, PLT clumps, lyse-resistant RBCs  cold agglutinins  Hct: extreme leukocytosis  microcytes, PLT clumps  extreme lipemia/leukocytosis
Interfering substances: differential	lyse-resistant RBCs, NRBCs, lipemia	—	NRBC, lyse-resistant RBCs, extreme hyperbilirubinemia
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed	60/60; 80/80 30 µL for CBC/30 µL/varies by tube size; 53 µL for CBC differential/53 µL for CBC differential/varies by tube size	—/35 differentials per hour —	60/60 30 µL for CBC and 53 µL for CBC and differential/ 30 µL for CBC and 53 µL for CBC and differential/—
Microsample capability Prepares microscope slides automatically or flags problems for slide prep	yes no	— —	yes yes
No. of automatic slidemakers available/List price	—	—	—
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes 10,000 samples 10,000 samples 10,000 samples no yes user or vendor no yes yes	yes/no unlimited ~4,000 — no — — — — —	yes/yes, with MultiLink Data Manager 100,000 unlimited with backup unlimited with backup yes yes user yes yes yes
LIS interface formats supported Information transferred on LIS interface	proprietary; proprietary ASTM numeric and flag results, histograms and differential plots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast	ASTM 1394 numeric and flag results, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast; host query for patient demographics and orders (when bar code is read, host is queried for orders)	ASTM 1394 and 1238, HL7, IEEE MIB numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, LIS to instrument—broadcast
LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube	no (yes on AL model) no Codabar, codes 39 and 128, Interleaved 2 of 5, EAN 8 and 13	no — Codabar, codes 39 and 128, Interleaved 2 of 5, QR, DataMatrix	yes no Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5
Accommodates bar-code placement per CLSI standard Auto2A	yes	—	yes
Time required for maintenance by lab personnel	none	less than 5 minutes per week	weekly: 15 minutes
Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	yes/no no	yes/no no	yes/yes yes, with Data Manager
Distinguishing features (supplied by company)	quantitative five-part WBC differential; aspirates only 30 µL of sample; requires small space footprint and runs quietly; AL has auto repeat based on decision rules	fully automated slide handling and oiling available in two models for medium and large laboratories; performs peripheral blood and body fluid differentials; WBC and other nucleated cells preclassified into 18 different categories; RBC morphology pre-characterized for 6 categories; network use allows remote review of blood smears and capability to link multiple analyzers in multiple locations, regardless of model	reliable five-part WBC differential technology—MTBF more than 200 days; small footprint; small sample size of 53 µL
Note: a dash in lieu of an answer means company did not answer question or question is not applicable			

## Hematology analyzers

Part 6 of 10	HORIBA Medical Jim Knowles jimknowles@horiba.com 34 Bunsen Irvine, CA 92618 888-903-5001 ext. 4553 www.horiba.com/us/en/medical	HORIBA Medical Jim Knowles jimknowles@horiba.com 34 Bunsen Irvine, CA 92618 888-903-5001 ext. 4553 www.horiba.com/us/en/medical	Siemens Healthcare Diagnostics Rita White rita.f.white@siemens.com 500 GBC Drive Newark, DE 19702 888-899-2896 www.usa.siemens.com/diagnostics
<b>Name of instrument</b> First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	<b>Pentra XL 80</b> 2004/2003/31 >250/>900/\$76,808	<b>Pentra DX120</b> 2005/2004/6 >20/>400/\$207,560	<b>Advia 120 Hematology System</b> 1998/1998/— >750/3,500/\$169,000–\$189,000
<b>Test menu:</b> • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, % neut, mono, lymph, eos, baso):  • Laboratory  • Flags	standard menu (left) plus: automatic dilution of overrange results (WBC × 3, RBC/hgb/PLT × 2), RDW, MPV  atypical lymphocytes, atypical lymphocytes %, LIC, LIC%  operator-selectable flagging	standard menu (left) plus: NRBCs, reticulocytes, IRF, MRV  LIC%&#, atypical lymphocytes %&#, IMG %&#, IML %&#, IMM %&#, RETL%, RETM%, RETH%, IMR%, MRU, MFI%, CRC%	standard menu (left) plus: CHCM, MPV, RDW, HDW, LUC %&#, retic %&#, CHR, CHCMr, MCVr; CSF: WBC, RBC, PMN, MN, neut, lymph, mono; cellular Hgb  %: hypo, hyper, macro, micro; calc. Hb, MPXI; %: blasts, PMN, MN; large PLT count; RBC fragment count; RBC ghost count; CSF: WBC, RBC, three-part differential; body fluids: TNC, RBC left shift, atyp. lymph, blasts, immature grans, myeloperoxidase deficiency, aniso, micro, macro, Hb variation, hypo, hyper, NRBC, RBC fragments, RBC ghost, large PLT, PLT clumps
FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only	— — — PCT, PDW, ATL, LIC	— — — PCT, PDW, ATL, LIC, IMG, IML, IMM	— — — IRF, MPC, MPM CSF, eos
Tests unique to analyzer	automatic dilution protocol	—	CHCM, HDW, CHR, CHCMr, MPC, MPM; CSF: WBC RBC, MN, PMN, neut, lymph, mono
<b>Differential method(s) used</b>	DHSS technology combining cytochemistry, focused flow impedance, and light absorbance	cytochemistry (chlorazol black E) and absorbance	perox–peroxidase cytochemistry staining with light scatter and absorption; baso–cytochemistry stripping with two-angle laser light scatter
<b>Linearity:</b> • WBC count/RBC count  • Hemoglobin/platelet • MCV (fL) or Hct (%)	0–120/0–8  0–24/0–1,900 (>2 g/dL Hb) 0–67 (Hct)/0–2,800 (<2 g/dL Hb)	0–150/0.5–8.1  2–25/0–2,000 0–80 (Hct)	0.02–400/0–7.0; CSF WBC 0–5,000/μL; CSF RBC 0–1,500/μL 0–22.5 /5–3,500 30–180 (MCV)
<b>Precision:</b> • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct	<2 percent/<2 percent <1 percent/<5 percent <2 percent (Hct)	<2 percent/<2 percent <1 percent/<5 percent <2 percent (Hct)	2.7 percent/1.2 percent 0.93 percent/2.93 percent 0.78 percent (MCV)
<b>Accuracy of automated differential compared with manual differential (per CLSI H-20A)</b>	neut% r=0.99, —; lymph% r=0.98, —; mono% r=0.96, —; eos% r=0.89, —; baso% r=0.54, —	neut% r=0.99, —; lymph% r=0.98, —; mono% r=0.92, —; eos% r=0.97, —; baso% r=0.71, —	neut% r=0.997, y=1.02x–0.6; lymph% r=0.997, y=1.00x+0.8; mono% r=0.943, y=0.85x–0.3; eos% r=0.979, y=0.87x+0.2; baso% r=0.772, y=0.67x+0.0; luc% r=0.994, y=0.92x+0.6 incomplete RBC lysis (perox only)
<b>Interfering substances:</b> • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	NRBCs, PLT clumps, lyse-resistant RBCs  cold agglutinins  Hct: extreme leukocytosis  microcytes, PLT clumps  extreme lipemia, leukocytosis	NRBCs, PLT clumps, lyse-resistant RBCs  cold agglutinins  Hct: extreme leukocytosis  microcytes, PLT clumps  extreme lipemia, leukocytosis	cold agglutinins, extreme sickle cell  none  none  high WBC, lip., extremely high bilirubin, interfere with cyanmethemoglobin only, none with direct cellular Hb (CHCM) incomplete lysis of RBCs, complete myeloperoxidase deficiency
<b>Interfering substances: differential</b>	NRBCs, lyse-resistant RBCs, extreme hyperbilirubinemia	NRBCs, lyse-resistant RBCs, extreme hyperbilirubinemia	
<b>Maximum CBCs per hour/Maximum CBCs and differentials per hour</b> <b>Minimum specimen volume open/Closed/Sample dead volume closed</b>	80/80 30 μL for CBC/53 μL for CBC and differential/0.5 mL	120/120 130 μL/200 μL/1 mL	120/120 157 μL/157 μL/<300 μL (tube size dependent)
<b>Microsample capability</b> Prepares microscope slides automatically or flags problems for slide prep	yes yes	yes, open mode yes	yes yes
<b>No. of automatic slidemakers available/List price</b>	—/—	—/—	—
<b>Archives patient data/Previous patient results incl. with recent results</b> <b>Maximum archived data accessible when system online</b> <b>No. specimens for which numeric results saved in memory at once</b> <b>No. specimens for which histo/cytogram results saved in memory at once</b> <b>Performs delta checks</b> <b>Tags and holds results for followup, confirmatory testing, or rerun</b> <b>Parameters for flags for holding samples defined by user or vendor</b> <b>Scattergram display: cell-specific color</b> <b>Histogram display: color with thresholds</b> <b>User interface can display choice of specimen/result information</b>	yes/yes, with MultiLink Data Manager 100,000 unlimited with backup unlimited with backup yes yes user yes yes yes —	yes/yes, with MultiLink Data Manager 100,000 unlimited with backup unlimited with backup yes yes user yes yes yes yes	yes/no 10,000 samples 10,000 samples 10,000 samples yes yes user or vendor yes yes yes
<b>LIS interface formats supported</b> <b>Information transferred on LIS interface</b>	proprietary, ASTM 1394 and 1238, HL7, IEEE MIB numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument— broadcast	proprietary, ASTM 1394 and 1238, HL7, IEEE MIB numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument— broadcast	proprietary (Spec 79) numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument— broadcast; host query for demographics and orders
<b>LOINC codes transmitted with all results</b> <b>Interface available or planned to automated specimen-handling system</b> <b>Bar-code symbologies read on specimen tube</b> <b>Accommodates bar-code placement per CLSI standard Auto2A</b>	— yes Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5 yes	— yes Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5 yes	no LabCell (Siemens) Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5 yes
<b>Time required for maintenance by lab personnel</b>	weekly: 15 minutes	weekly: 15 minutes	daily: 10 minutes; weekly: 15 minutes; monthly: 15 minutes
<b>Onboard diagnostics for troubleshooting/Limited to software problems</b> <b>Manufacturer can perform diagnostics via modem</b>	no/yes yes	no/yes yes	yes/no yes
<b>Distinguishing features (supplied by company)</b>	compact five-part differential instrument with autoloader and autodilution capability, auto rerun feature, autovalidation	high-throughput cell counter with integrated reticulocyte methodology and slidemaker/stainer; fluorescent NRBC counting, auto rerun and reflex testing, autovalidation	laser technology provides cellular Hb for RBCs and retics; 2-D PLT analysis that eliminates interference from RBC fragments and inclusion of large PLTs; dual WBC counts with a linearity of up to 400,000; CSF assay

Note: a dash in lieu of an answer means company did not answer question or question is not applicable

## Hematology analyzers

Part 7 of 10	Siemens Healthcare Diagnostics Rita White rita.f.white@siemens.com 500 GBC Drive Newark, DE 19702 888-899-2896 www.usa.siemens.com/diagnostics	Siemens Healthcare Diagnostics Rita White rita.f.white@siemens.com 500 GBC Drive Newark, DE 19702 888-899-2896 www.usa.siemens.com/diagnostics	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa
Name of instrument First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	Advia 2120 Hematology System 2004/2004/— >200/>900/\$225,000	Advia 2120i 2008/2008/130 >150/>400/\$225,000	Sysmex poch-100i 2004/2003/100 >950/>4,000/\$18,000
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, mono, lymph, eos, baso):  • Laboratory  • Flags  FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only Tests unique to analyzer	standard menu (left) plus: CHCM, MPV, RDW, HDW, LUC %&#, retic %&#, Chr, CHCMr, cellular Hgb, MCVr; CSF: WBC, RBC, PMN, MN, neut, lymph, mono % hypo, hyper, macro, micro; MPXI, %: blast, PMN, MN, large PLT count, RBC fragment count; RBC ghost count; NRBC; CSF: WBC, RBC, three-part differential; body fluids: TNC, RBC left shift, atyp. lymph, blasts, immature grans, myeloperoxidase deficiency, aniso, micro, macro, Hb variation, hypo, hyper, NRBC, RBC fragments, RBC ghost, large PLT, PLT clumps — — MPC, MPM IRF, CSF, eos CHCM, HDW, Chr, CHCMr, cellular Hgb, MPC, MPM, CSF: WBC, RBC, PMN, MN, neut, lymph, mono	standard menu (left) plus: CHCM, MPV, RDW, HDW, LUC %&#, retic %&#, Chr, CHCMr, cellular Hgb, MCVr; CSF: WBC, RBC, PMN, MN, neut, lymph, mono %hypo, hyper, macro, micro, MPXI, %blast, PMN, MN, large PLT count, RBC fragment count, RBC ghost count, NRBC; CSF: WBC, RBC, three-part differential; body fluids: TNC, RBC left shift, atyp. lymph, blasts, immature grans, myeloperoxidase deficiency, aniso, micro, macro, Hgb variation, hypo, hyper, NRBC, RBC fragments, RBC ghost, large PLT, PLT clumps — — MPC, MPM IRF, CSF eos CHCM, HDW, Chr, CHCMr, cellular Hgb, MPC, MPM, CSF: WBC, RBC, PMN, MN, neut, lymph, mono	WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, lymph, MXD, RDW-SD, RDW-CV, MPV —  histogram error flags; WBC, RBC, PLT — — — —
Differential method(s) used	peroxidase WBC—peroxidase cytochem. staining with light scatter and absorption; baso—cytochem. stripping with two-angle laser light scatter	peroxidase WBC: peroxidase cytochem. staining with light scatter and absorption; baso: cytochem. stripping with two-angle laser light scatter	direct current (DC)
Linearity: • WBC count/RBC count	0.02–400; CSF WBC 0–5,000/0–7.0; CSF RBC 0–1,500	0.02–400 CSF: 0–5,000/0–7.0 CSF: 0–1,500	1.0–99.9/0.3–7.0
Precision: • Hemoglobin/platelet • MCV (fL) or Hct (%) • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct	0–22.5/5–3,500 30–180 (MCV) 2.7 percent/1.2 percent 0.93 percent/2.93 percent 0.78 percent (MCV)	0–22.5/5–3,500 30–180 (MCV) 2.7 percent/1.2 percent 0.93 percent/2.93 percent 0.78 percent (MCV)	0.1–25.0/10–999 10–60 Hct <=3.5 percent/<=2.0 percent <=1.5 percent/<=6.0 percent <=2.0 percent Hct
Accuracy of automated differential compared with manual differential (per CLSI H-20A)	neut% r=0.997, y=1.02x–0.6; lymph% r=0.997, y=1.00x+0.8; mono% r=0.943, y=0.85x–0.3; eos% r=0.979, y=0.87x+0.2; baso% r=0.772, y=0.67x+0.0; luc% r=0.994, y=0.92x+0.6	neut% r=0.997, y=1.02x–0.6; lymph% r=0.997, y=1.00x+0.8; mono% r=0.943, y=0.85x–0.3; eos% r=0.979, y=0.87x+0.2; baso% r=0.772, y=0.67x+0.0; luc% r=0.994, y=0.92x+0.6	neut% R=0.98, LYM% R=0.99, MXD % R=0.75, neut# R=1.00, LYM# R=1.00, MXD# R=0.90
Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	incomplete RBC lysis (peroxidase only)  cold agglutinins, extreme sickle cell  —  —  extreme lipemia, high WBC, extreme high bilirubin interference with colorimetric Hb only, none with cellular Hb incomplete RBC lysis, complete myeloperoxidase deficiency	incomplete RBC lysis (peroxidase only)  cold agglutinins, extreme sickle cell  none  none  extreme lipemia, high WBC, extreme high bilirubin—interference with colorimetric Hgb only, none with cellular Hgb incomplete RBC lysis, complete myeloperoxidase deficiency	lyse-resistant RBC, cold agglutinins/cryoglobulins, PLT aggregation, NRBC cold agglutinins, microcytosis (severe), fragmented RBCs cold agglutinins, fragmented RBCs, leukocytosis (>100,000/uL) PLT aggregation, giant PLTs, microcytic RBCs, fragmented RBCs lipemia (severe), abnormal protein, leukocytosis (>100,000/uL)
Interfering substances: differential	—	—	—
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed	120/120 175 µL/175 µL/<300 (tube size dependent)	120/120 175 µL/175 µL/<300 (tube size dependent)	30/30 15 µL/15 µL/15 µL
Microsample capability Prepares microscope slides automatically or flags problems for slide prep	yes if integrated to Advia Autoslide	yes yes	yes no
No. of automatic slidemakers available/List price	Advia Autoslide, —/\$98,000	Advia Autoslide, —/\$98,000	—
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes/no 10,000 10,000 10,000 yes yes user or vendor yes yes yes	yes/no 10,000 samples 10,000 samples 10,000 samples yes yes yes yes yes yes	yes/yes 100 samples 100 samples 100 samples yes no yes no yes yes
LIS interface formats supported Information transferred on LIS interface	proprietary numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, patient orders, LIS to instrument—broadcast; host query for patient demographics and orders (when bar code is read, host is queried for orders)	proprietary (instrument or vendor specific) numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast; host query for demographics and orders	RS-232C numeric and flag results, histograms and scatterplots, patient demographics, patient orders, host query for patient demographics and orders
LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube	no LabCell (Siemens) Codabar, codes 39 and 128, Interleaved 2 of 5	no LabCell (Siemens) Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5	yes — codes 39 and 128, ASTM, ITF, NW7, JAN-8 and 13
Accommodates bar-code placement per CLSI standard Auto2A	—	yes	yes
Time required for maintenance by lab personnel	weekly: 15 minutes; monthly: 15 minutes	daily: 10 minutes; weekly: 15 minutes; monthly: 15 minutes	daily: <two minutes; weekly: <two minutes; monthly: <two minutes
Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	yes/no yes	yes/no yes	no/no yes
Distinguishing features (supplied by company)	laser technology provides direct cellular Hb for RBCs and reticulocytes; 2-D PLT analysis that eliminates interference from RBC fragments and inclusion of large PLTs; dual WBC counts with a linearity of up to 400,000; CSF assay	laser technology provides direct cellular Hgb for RBCs and reticulocytes; 2-D PLT analysis eliminates interference from RBC fragments and inclusion of large PLTs; dual WBC counts with a linearity of up to 400,000; CSF assay	hydrodynamic focusing, automatic floating discriminators, ISBT-compliant, data masking software for blood donor centers

Note: a dash in lieu of an answer means company did not answer question or question is not applicable

## Hematology analyzers

Part 8 of 10	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa
Name of instrument	Sysmex KX-21N	XS-1000i and XS-1000i AutoLoader (20 sample autoloader option)	Sysmex XT-1800i
First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	2001/1999/130 >1,100/>4,400/\$26,780	2006/2005/>320 >1,400/>6,000/\$85,000 (XS-1000i) \$95,000 (AutoLoader)	2002/2001/>40 >400/4,600/\$128,750
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, mono, lymph, eos, baso):  • Laboratory • Flags	WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, lymph, MXD, RDW-SD, RDW-CV, MPV  — histogram error flags; WBC, RBC, PLT	standard menu (left) plus: MPV, RDW-SD, RDW-CV  — PLT clumps, PLT abnormal distribution, WBC abnormal scattergram, blast immature granulocytes, left shift, atypical lymphocytes, abnormal lymphocytes/blasts, RBC abnormal distribution, RBC lyse resistance, RBC agglutinins, turbidity, NRBC  — IG% research screen  —	standard menu (left) plus: MPV, RDW-SD, RDW-CV, immature granulocytes %&#  — PLT clumps, PLT abnormal distribution, WBC abnormal scattergram, blast immature granulocytes, left shift, atypical lymphocytes, abnormal lymphocytes/blasts, RBC abnormal distribution, RBC lyse resistance, RBC agglutinins, turbidity, NRBC, body fluids  — — — immature granulocytes (IG%&#)
FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only Tests unique to analyzer	— — — — —	— — IG% research screen —	— — — — immature granulocytes (IG%&#)
Differential method(s) used	direct current (DC)	fluorescent flow cytometry	fluorescent flow cytometry
Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%)	1.0–99.9/0.3–7.0 0.1–25.0/10–999 10–60 Hct	0–400/0–8 0–25/0–5,000 0–60 (Hct)	0–310/0–8 0–25/0–5,000 0–60 (Hct)
Precision: • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct	<=3.5 percent/<=2.0 percent <=1.5 percent/<=6.0 percent <=2.0 percent Hct	—/— —/— —	<=3.0 percent/<=1.5 percent <=1.5 percent/<=4.0 percent <=1.5 percent (Hct)
Accuracy of automated differential compared with manual differential (per CLSI H-20A)	neut% R=0.98, LYM% R=0.99, MXD % R=0.75, neut# R=1.00, LYM# R=1.00, MXD# R=0.90	neut% r=0.96, y=0.9074x+3.8948; lymph% r=0.97, y=0.9017x+2.4817; mono% r=0.78, y=0.8626x+3.5938; eos% r=0.94, y=0.9076x+0.3651; baso% r=0.29, y=0.1538x+0.298	neut% r=0.95, y=0.95x+3.38; lymph% r=0.96, y=0.85x+1.67; mono% r=0.90, y=1.137x+1.89; eos% r=0.94, y=0.87x+0.04; baso% r=0.76, y=0.48x+0.24
Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	cold agglutinins, PLT aggregation, erythroblastosis, NRBC, cryoglobulins cold agglutinins, severe microcytosis, fragmented RBCs, leukocytosis (>100,000/ $\mu$ L)  Hct: cold agglutinin, leukocytosis (>100,000/ $\mu$ L), abnormal red cell fragility, spherocytosis  pseudothrombocytopenia, PLT aggregation, increased microcytosis, megalocytic PLTs  leukocytosis (>100,000/ $\mu$ L), lipemia, abnormal protein	cold agglutinins, PLT aggregation, cryoglobulins, lyse-resistant RBCs, NRBCs cold agglutinins, severe microcytosis, fragmented RBCs, leukocytosis  Hct: cold agglutinins, abnormal red cell fragility, spherocytosis, leukocytosis (>100,000/ $\mu$ L)  pseudothrombocytopenia, PLT aggregation, increased microcytosis, megaloblasts  lipemia, abnormal proteins, leukocytosis (>100,000/ $\mu$ L) lyse-resistant RBCs	cold agglutinins, PLT aggregation, cryoglobulins, lyse-resistant RBCs, NRBCs cold agglutinins, severe microcytosis, fragmented RBCs, leukocytosis  Hct: cold agglutinins, abnormal red cell fragility, spherocytosis, leukocytosis (>100,000/ $\mu$ L)  pseudothrombocytopenia, PLT aggregations, increased microcytosis, megaloblasts  lipemia, abnormal proteins, leukocytosis (>100,000/ $\mu$ L) lyse-resistant RBCs
Interfering substances: differential	—	—	—
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed	60/60 50 $\mu$ L/—/—	60/60 20 $\mu$ L/20 $\mu$ L/1.0 mL	80/80 85 $\mu$ L/150 $\mu$ L/1 mL
Microsample capability Prepares microscope slides automatically or flags problems for slide prep	yes no	yes no	yes no
No. of automatic slidemakers available/List price	—	—	—
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes/yes 300 samples 300 samples 300 samples yes yes yes no yes yes	yes/yes 10,000 samples 10,000 samples 10,000 samples yes yes user or vendor yes yes yes	yes/yes 10,000 samples 10,000 samples 10,000 samples yes yes user or vendor yes yes yes
LIS interface formats supported Information transferred on LIS interface	RS-232C numeric and flag results, histograms and scatterplots, host query for patient demographics and orders	proprietary, ASTM 1394, TCP-IP numeric and flag results, histograms and scatterplots, patient demographics, orders	RS-232C/TCP-IP, ASTM numeric and flag results, histograms and scatterplots, patient demographics, orders
LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube	yes — codes 39 and 128, ITF, NW-7, JAN, UPC-A, UPC-E, EAN13, EAN8	yes — Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5, NW7, EAN 8 and 13, ITF	yes — Codabar, codes 39 and 128, Interleaved 2 of 5, ITF, NW7, EAN 8 and 13
Accommodates bar-code placement per CLSI standard Auto2A	yes	yes	yes
Time required for maintenance by lab personnel	daily: <two minutes; weekly: <two minutes; monthly: <two minutes	daily: three minutes; weekly: none; monthly: nine minutes	daily: <three minutes
Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	no/no yes	yes/no yes, also via Internet	yes/no yes, also via Internet
Distinguishing features (supplied by company)	automatic floating discriminators	standardized technology, reagents, controls, and operations with other X series analyzers; small sample volume requirements for CBC and five-part differential; remote diagnostics, online QC, discrete analysis, reagent monitoring, chartable report; remote calibration verification	remote diagnostics; online QC; random access; discrete testing; reagent monitoring; chartable report formats; unique specimen-gating, software is FDA Part II compliant; body fluids now FDA cleared; standardized technology, reagents, controls, and operations with other X series analyzers; XT-V for use in toxicology, research, and veterinary reference labs

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## Hematology analyzers

Part 9 of 10	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa
Name of instrument First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	Sysmex XT-2000i 2002/2001/>70 >900/>5,200/\$149,500	Sysmex XT-4000i 2010/2009/80 100/>250/\$195,700	Sysmex XE-2100D 2004/2004/>15 190/>205/\$200,000
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, mono, lymph, eos, baso):  • Laboratory • Flags	standard menu (left) plus: retic %&#, IRF, PLT-O, MPV, RDW-SD, RDW-CV, reticulocyte hemoglobin, immature granulocytes %&#  — PLT clumps, PLT abnormal distribution, WBC abnormal scattergram, blast immature granulocytes, left shift, atypical lymphocytes, abnormal lymph/blasts, RBC abnormal distribution, RBC lyse resistance, RBC agglutinins, turbidity, NRBC, body fluids  — — — — PLT-O, immature granulocytes (IG) %&#, reticulocyte hemoglobin (RET-He)	standard menu (left) plus: IG% and #, retic % and #, IRF, RET-He, PLT-O, BF: RBC/WBC/TC/two-part differential  — PLT clumps, PLT abnormal distribution, blast immature granulocytes, left shift, atypical lymphocytes, abnormal lymph/blasts, NRBC, RBC lyse resistance, RBC abnormal distribution, RBC agglutination, turbidity  — — — — reticulocyte hemoglobin, immature reticulocyte fraction, reportable immature granulocyte # and %, PLT-O, BF: RBC/WBC/TC/two-part differential	standard menu (left) plus: RDW-SD, RDW-CV  — PLT clumps, PLT abnormal distribution, WBC abnormal scattergram, blast, left shift, atypical lymphocytes, abnormal lymph/blasts, RBC abnormal distribution, RBC lyse resistance, RBC agglutinins, turbidity  — — — P-LCR, PCT, PDW optional: IG% & IG#
Differential method(s) used Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%) Precision: • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct Accuracy of automated differential compared with manual differential (per CLSI H-20A)	fluorescent flow cytometry 0-310/0-8 0-25/0-5,000 0-60 (Hct) ≤3.0 percent/≤1.5 percent ≤1.5 percent/≤4.0 percent ≤1.5 percent (Hct) neut% r=0.95, y=0.95x+3.38; lymph% r=0.96, y=0.85x+1.67; mono% r=0.90, y=11.37x+1.89; eos% r=0.94, y=0.87x+0.04; baso% r=0.76, y=0.48x+0.24	fluorescent flow cytometry 0-440/0-8 0-25/0-5,000 0-60 (Hct) ≤3.0 percent/≤1.5 percent ≤1.5 percent/≤4.0 percent ≤1.5 percent (Hct) neut% r=0.95, lymph% r=0.96, mono% r=0.90, eos% r=0.94, baso% r=0.76; neut % y=0.95x+3.38, lymph % y=0.85x+1.67, mono % y=11.37x+1.89, eos% y=0.87x+0.04, baso% y=0.48x+0.24	fluorescent flow cytometry 0-440/0-8 0-25/0-5,000 0-75 (Hct) ≤3 percent/≤1.5 percent ≤1.0 percent/≤4.0 percent ≤1.5 percent (Hct) neut% r=0.95, y=0.92x+5.46; lymph% r=0.95, y=0.88x+2.46; mono% r=0.79, y=0.77x+1.88; eos% r=0.92, y=0.97x+0.29; baso% r=0.82, y=1.01x+0.01; NRBC% r=0.96, y=1.12x+0.11; IG% r=0.83, y=0.9332x+0.0922
Interfering substances: • WBC  • RBC  • MCV or Hct  • Platelet  • Hemoglobin	cold agglutinins, PLT aggregation, cryoglobulins, lyse-resistant RBCs, NRBCs  cold agglutinins, severe microcytosis, fragmented RBCs, leukocytosis  Hct: cold agglutinins, abnormal red cell fragility, spherocytosis, leukocytosis (>100,000/μL)  pseudothrombocytopenia, PLT aggregation, increased microcytosis, megaloblasts  lipemia, abnormal proteins, leukocytosis (>100,000/μL)	cold agglutinins, PLT aggregation, cryoglobulin, lyse-resistant erythrocytes, NRBC  cold agglutinins, severe microcytosis, fragmented RBCs, leukocytosis  Hct: cold agglutinins, fragmented RBCs, spherocytosis, leukocytosis (lymphocytes>100,000/μL)  PLT aggregation, pseudothrombocytopenia, giant platelets, microcytosis, cryoglobulin  leukocytosis (lymphocytes>100,000/μL), lipemia, abnormal protein	cold agglutinins, PLT aggregation, cryoglobulin, lyse-resistant RBCs, NRBCs  cold agglutinins, severe microcytosis, fragmented RBCs, leukocytosis  Hct: cold agglutinins, abnormal red cell fragility, spherocytosis, leukocytosis  pseudothrombocytopenia, PLT aggregation, increased microcytosis, megaloblasts  lipemia, abnormal proteins, leukocytosis (>100,000/μL)
Interfering substances: differential	lyse-resistant RBCs	lyse-resistant RBCs	lyse-resistant RBCs
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed	80/80 85 μL/150 μL/1 mL	100/100 85 μL/150 μL/1 mL	150/150 130 μL/200 μL/1 mL
Microsample capability Prepares microscope slides automatically or flags problems for slide prep No. of automatic slidemakers available/List price	yes no —	yes no	yes yes, with Alpha or HST upgrade >1,000/price depends on configuration
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once	yes/yes 10,000 samples 10,000 samples 10,000 samples	yes/yes 10,000 samples 10,000 samples 10,000 samples	yes/yes 10,000 samples 10,000 samples 10,000 samples
Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes yes user or vendor yes yes yes	yes yes yes yes yes yes	yes yes user or vendor yes yes yes
LIS interface formats supported Information transferred on LIS interface	RS-232/TCP-IP, ASTM numeric and flag results, histograms and scatterplots, patient demographics, orders	ASTM numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast; host query for demographics and orders	RS-232C/TCP IP numeric and flag results, histograms and scatterplots, patient demographics, orders
LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube	yes — Codabar, codes 39 and 128, Interleaved 2 of 5, ITF, NW7, EAN 8 and 13	yes — Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5, ITF, NW7	yes on automation platform Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5, ITF, NW7, EAN 8 and 13, ISBT
Accommodates bar-code placement per CLSI standard Auto2A	yes	yes	yes
Time required for maintenance by lab personnel	daily: <three minutes	daily: <three minutes	daily: <three minutes
Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	yes/no yes, also via Internet	yes/no yes	yes/no yes, also via Internet
Distinguishing features (supplied by company)	high throughput, remote diagnostics; online QC; random access; fluorescent optical platelets; discrete testing; reagent monitoring; customized chartable report formats; body fluids, standardized technology, reagents, controls, and operations with other X series analyzers; IG # & %, RET-He; XT-V unit for use in toxicology, research, and veterinary reference labs	testing parameters: fluorescent optical platelets, IG #&%, RET-He, body fluids (CSF, serous, synovial), WBC/RBC/TC and two-part differential; standardized technology, reagents, controls, and operations with other X series analyzers; simplified operations with extended linearities, high-throughput, remote-monitoring capabilities	150 CBCs per hour; platelet linearity—5 million, hematocrit extended to 75 percent; standardized technology, reagents, controls and operations; ISBT-compliant; FDA-cleared application for blood component products in specified anticoagulants

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## Hematology analyzers

Part 10 of 10	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa	Sysmex America Tammy Kutz communications@sysmex.com 1 Nelson C. White Parkway Mundelein, IL 60060 800-379-7639 www.sysmex.com/usa
Name of instrument	Sysmex XE-2100	Sysmex XE-5000	Sysmex XE-Alpha N/HST-N
First year installed in U.S./Outside U.S./No. of units sold in 2010 No. units installed in U.S./Outside U.S./List price	1999/—/~75 1,325/>5,000/\$240,000	2008/2008/>125 >640/>2,000/\$265,122	2000/—/>120 >950/1,400/\$360,000–\$1,000,000
Test menu: • Chartable (standard menu: WBC, RBC, Hb, Hct, MCV, MCH, MCHC, PLT, %&# neut, mono, lymph, eos, baso):  • Laboratory • Flags  FDA-cleared tests not clinically released Tests not available but submitted for 510(k) clearance Tests in development Tests for research use only Tests unique to analyzer	standard menu (left) plus: NRBC %&#, retic %&#, RDW-SD, RDW-CV, IRF, PLT-O, HPC#, MPV, IG%, IG#, RET-He, IPF — PLT clumps, RBC agglut, turbidity, WBC abnormal scattergram, RBC abnormal distribution, PLT abnormal distribution, RBC lyse resistance, blasts, left shift, atypical lymphocytes, abnormal lymph./blast, reticulocyte abnormal scattergram — — P-LCR, PCT, PDW HPC#, IG%, IG#, RET He, IPF	standard menu (left) plus: NRBC %&#, retic %&, RDW-SD, RDW-CV, IRF, PLT-O, HPC#, MPV, IG%, IG#, RET-He, IPF — PLT clumps, PLT abnormal distribution, WBC abnormal scattergram, blast, left shift, atypical lymphocytes, abnormal lymph./blast, RBC abnormal distribution, RBC lyse resistance, RBC agglut., turbidity — — — reticulocyte hemoglobin, immature platelet fraction, hematopoietic progenitor cell, immature reticulocyte fraction, reportable immature granulocyte #&%, RBC/WBC/TC/two-part differential	standard menu (left) plus: NRBC%&, retic%&, RDW-SD, RDW-CV, IRF, PLT-O, HPC#, MPV, IG%, IG#, RET-He, IPF — user-defined, all-inclusive — — — P-LCR, PCT, PDW NRBC, HPC#, IG%, IG#, RET-He, immature platelet function (IPF)
Differential method(s) used	fluorescent flow cytometry, RF/DC detecting method	fluorescent flow cytometry, RF/DC detection method	fluorescent flow cytometry, RF/DC detecting method
Linearity: • WBC count/RBC count • Hemoglobin/platelet • MCV (fL) or Hct (%) Precision: • WBC count/RBC count • Hemoglobin/platelet • MCV or Hct Accuracy of automated differential compared with manual differential (per CLSI H-20A)	0–440/0–8 0–25/0–5,000 0–75 (Hct) <3 percent/<1.5 percent <1.0 percent/<4.0 percent <1.5 percent (Hct) neut% r=0.95, y=0.92x+5.46; lymph% r=0.95, y=0.88x+2.46; mono% r=0.79, y=0.77x+1.88; eos% r=0.92, y=0.97x+0.29; baso% r=0.82, y=1.01x+0.01; NRBC% r=0.96, y=1.12x+0.11; IG% r=0.83, y=0.9332x+0.0922	0–440/0–8 0–25/0–5,000 0–75 (Hct) <3 percent/<1.5 percent <1.0 percent/<4.0 percent <1.5 percent (Hct) neut% r=0.95, y=0.92x+5.46; lymph% r=0.95, y=0.88x+2.46; mono% r=0.79, y=0.77x+1.88; eos% r=0.92, y=0.97x+0.29; baso% r=0.82, y=1.01x+0.01; NRBC% r=0.96, y=1.12x+0.11; IG% r=0.83, y=0.9332x+0.0922	0–440/0–8 0–25/0–5,000 0–75 (Hct) <3 percent/<1.5 percent <1.0 percent/<4.0 percent <1.0 percent (Hct) neut% r=0.95, y=0.92x+5.46; lymph% r=0.95, y=0.88x+2.46; mono% r=0.79, y=0.77x+1.88; eos% r=0.92, y=0.97x+0.29; baso% r=0.82, y=1.01x+0.01; NRBC% r=0.96, y=1.12x+0.11; IG% r=0.83, y=0.9332x+0.0922
Interfering substances: • WBC	cold agglutinins, PLT aggregation, nucleated RBCs, cryoglobulin, lyse-resistant RBCs	cold agglutinins, PLT aggregation, nucleated RBCs, cryoglobulin, lyse-resistant RBCs	cold agglutinins, PLT aggregation, nucleated RBCs, cryoglobulins, lyse-resistant RBCs
• RBC	cold agglutinins, severe microcytosis, fragmented RBCs, large No. giant PLTs, in vitro hemolysis	cold agglutinins, severe microcytosis, fragmented RBCs, large number giant PLTs, in vitro hemolysis	cold agglutinins, severe microcytosis, fragmented RBCs, large No. giant PLTs, in vitro hemolysis
• MCV or Hct	Hct: cold agglutinins, leukocytosis, abnormal red cell fragility, spherocytosis	Hct: cold agglutinins, leukocytosis, abnormal red cell fragility, spherocytosis	Hct: cold agglutinins, leukocytosis, abnormal red cell fragility, spherocytosis
• Platelet	pseudothrombocytopenia, PLT aggregation, increased microcytosis, megalocytic PLTs	pseudothrombocytopenia, PLT aggregation, increased microcytosis, megalocytic PLTs	pseudothrombocytopenia, PLT aggregation, increased microcytosis, megalocytic PLTs
• Hemoglobin	lipemia, abnormal proteins, leukocytosis (>100,000/ $\mu$ L) lyse-resistant RBCs	lipemia, abnormal proteins, leukocytosis (>100,000/ $\mu$ L) lyse-resistant RBCs	lipemia, abnormal proteins, leukocytosis (>100,000/ $\mu$ L) lyse-resistant RBCs
Interfering substances: differential			
Maximum CBCs per hour/Maximum CBCs and differentials per hour Minimum specimen volume open/Closed/Sample dead volume closed	150/150 130 $\mu$ L/200 $\mu$ L/1 mL	150/150 130 $\mu$ L/200 $\mu$ L/1 mL	150/150 per analyzer on automation system 130 $\mu$ L/200 $\mu$ L/1 mL
Microsample capability Prepares microscope slides automatically or flags problems for slide prep	yes yes (with Alpha or HST upgrade)	yes yes (with Alpha or HST upgrade)	yes yes
No. of automatic slidemakers available/List price	>1,000/price depends on configuration	>1,200/price depends on configuration	>1,700/\$250,000
Archives patient data/Previous patient results incl. with recent results Maximum archived data accessible when system online No. specimens for which numeric results saved in memory at once No. specimens for which histo/cytogram results saved in memory at once	yes/yes 10,000 samples 10,000 samples 10,000 samples	yes/yes 10,000 samples 10,000 samples 10,000 samples	yes/yes 10,000 samples 10,000 samples; 20,000 orders 10,000 samples; two years plus, with optional decision logic software
Performs delta checks Tags and holds results for followup, confirmatory testing, or rerun Parameters for flags for holding samples defined by user or vendor Scattergram display: cell-specific color Histogram display: color with thresholds User interface can display choice of specimen/result information	yes yes user or vendor yes yes yes yes	yes yes yes yes yes yes yes	yes yes user and vendor yes yes yes yes
LIS interface formats supported Information transferred on LIS interface	RS-232C/TCP IP numeric and flag results, histograms and scatterplots, patient demographics, orders	ASTM 1394, TCP-IP, ASTM E1381 numeric and flag results, histograms and scatterplots, instrument to LIS; patient demographics, orders, LIS to instrument—broadcast; host query for demographics and orders	RS-232C/TCP IP numeric and flag results, histograms and scatterplots, patient demographics, orders
LOINC codes transmitted with all results Interface available or planned to automated specimen-handling system Bar-code symbologies read on specimen tube	yes on automation platform Codabar, codes 39 and 128, Interleaved 2 of 5, ITF, NW7, EAN 8 and 13	yes Roche Diagnostics, and Labotix, A & T, Thermo, IDS Codabar, codes 39 and 128, ASTM, Interleaved 2 of 5, ITF, NW7, EAN 8 and 13	yes Roche, Labotix, IDS, A & T, Thermo engen Codabar, codes 39 and 128, Interleaved 2 of 5, ITF, NW7, EAN 8 and 13
Accommodates bar-code placement per CLSI standard Auto2A	yes	—	yes
Time required for maintenance by lab personnel	daily: <three minutes	daily: <three minutes	daily: <three minutes (operator time)
Onboard diagnostics for troubleshooting/Limited to software problems Manufacturer can perform diagnostics via modem	yes/no yes, also via Internet	yes/no yes, also via Internet	yes/no yes, also via Internet
Distinguishing features (supplied by company)	throughput of 150 CBCs per hour; random access; discrete testing; online QC; remote diagnostics, body fluid analysis; platelet linearity to 5 million, hematocrit linear to 75 percent; hematopoietic progenitor cell testing; immature granulocyte enumeration; immature platelet fraction; reticulocyte hemoglobin equivalent; standardized reagents, controls, and operations with other X series analyzers	low-end linearity for all body fluids; two-part differential (mono nuclear % + # and polymorphonuclear % + # or body fluid; reticulocyte hemoglobin content; immature platelet fractions; throughput of 150 CBCs per hour; random access; discrete testing; online QC; remote diagnostics, body fluid analysis; platelet linearity to 5 million, hematocrit linear to 75 percent; hematopoietic progenitor cell testing; immature granulocyte enumeration; immature platelet fraction; reticulocyte hemoglobin equivalent; standardized reagents, controls, and operations with other X series analyzers	high-throughput, flexible, scalable configurations available (>125 standard configurations available); platelet linearity—5 million; new parameters for platelet monitoring—IPF and reticulocyte Hb measurement and RET-He, hematopoietic progenitor cell analysis, lavender top management, standardized technology, reagents, controls, and operations; broader clinical reportable ranges; enhanced clinical parameters to support preventive care and disease management

Note: a dash in lieu of an answer means company did not answer question or question is not applicable