Q&A column

Editor: Frederick L. Kiechle, MD, PhD

Submit your pathology-related question for reply by appropriate medical consultants. CAP TODAY will make every effort to answer all relevant questions. However, those questions that are not of general interest may not receive a reply. For your question to be considered, you must include your name and address; this information will be omitted if your question is published in CAP TODAY.

Table 1				
HER2 data	First ISH count			
HER2 copies/cell	4.80			
Centromere 17 signals/cell	2.75			
Ratio of HER2/CEP17 signals	1.74			



Fig. 1. Dual-probe in situ hybridization: HER2=red, CEP17=green, showing increased HER2 signals.

Q. I just read the 2018 HER2 guideline update. Can you provide an example of how a previously equivocal case is resolved under the new guideline?

A. March 2019—The 2018 revision to the American Society of Clinical Oncology/College of American Pathologists HER2 testing in breast cancer guideline contains several timely updates and a number of flow charts relevant to nonclassic in situ hybridization categories, along with interpretive and managerial comments that can be

incorporated into pathology reporting.¹ Here are examples:

This ISH result would be in the equivocal category in the 2013 guideline, based on a *HER2*/CEP17 ratio of less than 2.0, in light of an average 4.8 *HER2* copies per cell.



Fig. 2. Corresponding HER2 immunohistochemistry (4B5 clone), showing moderate intensity circumferential and noncircumferential staining, interpreted as 2+.

In figures 3 and 6 in the 2018 guideline, this is a "Group 4" result, requiring correlation with HER2 immunohistochemistry.¹ HER2 IHC should be performed on the same block or sections used for ISH, preferably by the same institution, and interpreted in parallel (shown in **Fig. 2** for this case). Alternate chromosome 17 probes are no longer necessary and are discouraged.

If the IHC result is definitive, overall interpretation is based on that IHC result. In other words, if HER2 IHC is 0 or

1+, the overall interpretation is negative. If HER2 IHC is 3+, the overall interpretation is positive.¹ If HER2 IHC is 2+, as it so often is in cases sent for ISH, additional ISH cells should be counted by a second observer, blinded to the original ISH results, but with attention to the 2+ or *strongest staining areas* of the carcinoma (**Table 2** for this

example). If the result remains the same, the overall interpretation is HER2 negative, as delineated (Table 3).¹

The overall interpretation for a 2+ IHC result differs by ISH group, as summarized in Table 3 and fully detailed in

HER2 data	First ISH count	Second ISH count				
HER2 copies/cell	4.80	4.42				
Centromere 17 signals/cell	2.75	2.60				
Ratio of <i>HER2</i> / CEP17 signals	1.74	1.70				
HER2 immunohisto- chemistry	2+					
Overall interpretation	HER2 negative (based on ISH and IHC)					

the guideline.

fable 3. 2018 ASCO/CA	^o dual-probe HER2 I	ISH interpretation [†]
-----------------------	--------------------------------	---------------------------------

Group	HER2/CEP17 ratio	<i>HER2</i> signals/cell	Interpretation	Further workup and interpretation
1	≥2	≥4	ISH positive	
2	≥2	<4	Further workup	Correlate with IHC; if 2+, count additional ISH cells* HER2 NEGATIVE unless IHC 3+
3	<2	≥6	Further workup	Correlate with IHC; if 2+, count additional ISH cells* HER2 POSITIVE unless IHC 0 or 1+
4	<2	\geq 4 and < 6	Further workup	Correlate with IHC; if 2+, count additional ISH cells* HER2 NEGATIVE unless IHC 3+
5	<2	<4	ISH negative	

*If ISH result is different based on additional cell counts, results are adjudicated based on standard procedures and used for overall interpretation. †See guideline for additional details.¹

1. Wolff AC, Hammond MEH, Allison KH, et al. Human epidermal growth

factor receptor 2 testing in breast cancer: American Society of Clinical Oncology/College of American Pathologists clinical practice guideline focused update. *Arch Pathol Lab Med.* 2018;142(11):1364–1382.

Megan Troxell, MD, PhD Professor, Department of Pathology Stanford University School of Medicine, Stanford, Calif. Advisor, CAP Immunohistochemistry Committee

Andrew Michael Bellizzi, MD Clinical Associate Professor, Department of Pathology, University of Iowa Hospitals and Clinics Co-director of Immunopathology Laboratory, Director of GI Pathology, Holden Comprehensive Cancer Center, Iowa City, Iowa Chair, CAP Immunohistochemistry Committee